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Seasonal Variation in the Leaf Essential Oil Composition of Zanthoxylum clava-herculis growing in Huntsville, Alabama

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The seasonal variation in the chemical composition of the leaf essential oil of *Zanthoxylum clava-herculis* has been analyzed by GC-MS. Three individual trees were sampled four times during the course of the 2004 growing season. Notable differences were recorded in the essential oil yields from the three trees on the four collection dates; yields were highest in May and lowest in July The leaf essential oils were made up of 25 components, largely menthane monoterpenoids, dominated by limonene (44-73%) and 1,8-cineole (16-43%), with lesser amounts of α -thujene, linalool, γ -terpinene, and α -terpineol. The ratio of oxygenated monoterpenoids to monoterpene hydrocarbons generally increased during the season, largely reflected in the 1,8-cineole/limonene ratio.

Keywords: Zanthoxylum clava-herculis, Rutaceae, seasonal variation, essential oil, GC-MS, limonene, 1,8-cineole.

Zanthoxylum clava-herculis L. (Rutaceae; toothache tree. Southern prickly-ash. Hercules' club) is a small. deciduous tree growing to 10 m in height and 15 cm dbh. The flowers appear in April-May, and the fruits, which are small follicles that develop between July-September, dehisce to reveal shiny black seeds. The tree, a temperate representative of a circumtropical genus, ranges from Virginia south to Florida and west to Texas, Arkansas, and Oklahoma [1a]. The seeds are used as a condiment and a pepper substitute [1b]. The bark of Z. clava-herculis has been used by native Americans for chronic rheumatism, dyspepsia, dysentery, kidney trouble, heart trouble, colds, coughs, lung ailments, toothache, and nervous debility [1c,1d]. Bioactive principles include herculin and neoherculin (insecticidal), berberine and chelerythrine (antibacterial), N-acetylanonaine (ichthyotoxic), as well asarinin. sesamin. *O*-prenylpiperitol, as 0prenylxanthoxylol, and xanthoxylin [1d,2]. Z. clavaherculis is also a host plant of several insects, including swallowtail butterflies [3] and leaf beetles [4].

In this paper, we present the identity and seasonal variation in the chemical composition of the leaf essential oils of *Z. clava-herculis* growing in Huntsville, Alabama, during the growing season of 2004. To our knowledge, this report is the first to present the identity and seasonality of *Z. clava-herculis* leaf oil.

The yields from the collection and hydrodistillation of fresh leaves from *Z. clava-herculis* are listed in Table 1. The results show some notable differences in the essential oil yields from the three trees on the four collection dates. Yields for the three trees were highest in May and lowest in July. Among the three trees, the female one had a higher yield than the two male trees. Essential oil yields ranged from 1.1% (from the female tree in May) to 0.4% (from a male tree in July). The weather data for Huntsville, Alabama during the four collection dates in 2004 are shown in Table 2. Average temperatures were lowest in May and highest in July. Rainfall was highest in July and at least three-fold higher than in May. Average sunlight was also highest in July.

The gas chromatographic-mass spectral analyses are summarized in Table 3. A total of 25 components were detected in the leaf oils, but not all components were detected in the three plants or from the different collection dates. Monoterpene hydrocarbons and oxygenated monoterpenoids were dominant, with very small amounts of sesquiterpenes and simple oxygenated compounds (*cis*-3-hexenol and nonanal). The major components were limonene (44-73%) and 1,8-cineole (16-43%). Other components detected in relatively less amounts include α -thujene, linalool, γ -terpinene, and α -terpineol. These compounds were all detected in the

three plants at the four collection dates. Limonene has also been reported as an an abundant component of Z. *limonella* [5] and Z. *piperitum* [6] fruit essential oils, while 1,8-cineole is an abundant constituent of Z. *acuminatum* leaf essential oil [7]. Linalool and α -terpineol seem to be ubiquitous components of Zanthoxylum leaf oils [7]. α -Thujene and γ -terpinene are common constituents of leaf oils of Citrus and other Rutaceae [8].

Traa	Date Collected	Mass Fresh	Mass Ess.	Yield Ess.
filee	Extracted	Leaves (g)	Oil (g)	Oil (%)
	12-May-04	20.26	0.195	1.0
A (7)	3-Jul-04	24.57	0.104	0.4
A (0)	26-Aug-04	20.03	0.153	0.8
	23-Oct-04	25.86	0.158	0.6
	13-May-04	18.32	0.134	0.7
$\mathbf{P}(\mathcal{A})$	4-Jul-04	19.34	0.073	0.4
в(0)	27-Aug-04	15.76	0.132	0.8
	24-Oct-04	22.15	0.148	0.7
	14-May-04	15.85	0.181	1.1
C(0)	5-Jul-04	18.34	0.116	0.6
C (¥)	28-Aug-04	15.23	0.138	0.9
	24-Oct-04	21.27	0.151	0.7

 Table 1: Leaf essential oil yields of Zanthoxylum clava-herculis.

Certain components detected in the female tree were not detected in the male trees, including γ -muurolene, valencene, γ -cadinene, and δ -cadinene. Also, α -terpineol was detected in greater amounts in the female tree than in the two male trees. Most of the remaining components were either detected in trace amounts or undetected in some trees or dates. Comparing the abundance of the major components across the collection dates, limonene was most abundant in the May collections, whereas 1,8-cineole showed a gradual increase in abundance from May to October. A similar trend was recorded for all three trees.

It is not readily apparent what factors may be responsible for the variation in essential oil chemical compositions. *Salvia libanotica* from Lebanon shows a similar increase in 1,8-cineole concentration with concomitant decrease in limonene concentration in August compared with April (57.4% cineole and 2.4% limonene in August; 47.7% cineole and 6.1% limonene in April) [9]. Similar seasonal changes have been reported for *Pinus pinea* from both southern Italy, May vs. October [10], and Spain, early June vs. early September [11a]; and for *Picea sitchensis* during the growing season [11b].

The weather data for the two-week periods prior to collection, summarized in Table 2, may explain, at least in part, the seasonal variation in *Z. clava-herculis* leaf essential oil yields recorded in this study. It may be that the relatively warm temperatures, coupled with abundant sunshine and rainfall, during the two weeks prior to the early July collection was responsible for the lower essential oil yields at that time by increasing the

 Table 2: Weather data for Hunstville, Alabama during collection of plant materials in 2004.

	29-Apr to 12- May	20-Jun to 3- Jul	13-Aug to 26- Aug	8-Oct to 21- Oct
Av High (°C) ^a	26.7	27.9	28.6	21.2
Av Low (°C) ^a	13.1	20.4	16.6	12.9
Av Temp (°C) ^a	19.9	24.1	22.6	17.1
Rainfall (mm) ^b	40	146	23	122
Sunshine (hours) ^a	50	77	49	99
Av Daylight (hrs:min)	13:43	14:27	13:18	11:21
Tree phenology	flowering		fruiting	

^{*a*} National Weather Service Forecast Office, Huntsville, Alabama [12]. ^{*b*} Cooperative Huntsville Area Rainfall Measurements (Alabama) [13]

emission of volatiles and decreasing the amount of essential oil remaining in the leaves. Monoterpene emissions are generally maximized during the warmer part of the season. Thus, for example, monoterpene emissions for both *Pinus pinea* and *Quercus ilex* from Spain were observed in June, attributed to high temperatures and insolation [11a]. Similarly, monoterpene emissions from *Eucalyptus globulus* from Western Australia were greatest during the Australian summer (January) and lowest during the winter (July) [14].

Although the sample size was small and not statistically relevant, there seems to have been less chemical diversity (fewer components) in the two male trees sampled compared with the single female tree. The increased chemical diversity of the female tree was manifested in the sesquiterpenoids (γ -muurolene, valencene, γ -cadinene, and δ -cadinene) that were not detected in the male trees. In addition, with the exception of 1,8-cineole, the female tree had higher concentrations of oxygenated monoterpenoids (sabinene hydrates, linalool, α -terpineol, and terpinen-4-ol) and higher concentrations of the sesquiterpenoid Bcaryophyllene. In addition to higher chemical diversity, the female tree showed slightly higher essential oil yields than either of the two male trees. Higher essential oil yield and chemical diversity in female trees has been previously observed in Juniperus virginiana [15]. Similarly, the floral essential oils from female Laurus nobilis were more chemically diverse than that from males [16a] and Ficus carica female trees showed greater chemical diversity in their fig volatiles than male trees [16b]. Non-volatile bark constituents of the medicinal plant Amphipthervgium adstringens were present in greater quantities in female than male trees [17]. Female Cannabis sativa plants generally produced higher levels of total cannabinoids [18a], as well as terpenoids [18b], than male plants. Since sexual reproduction is generally thought to be metabolically more costly for female plants than for males [19], increased investment in essential oils in females relative to males might represent the product of sexually biased natural selection for compounds conferring some protection against herbivores and pathogens. It is also

Table 3: Seasonal variation in leaf essential oil composition of Zanthoxylum clava-herculis.

		Area (%)											
RI ^a Compound		Tree A (d)			Tree B (d)			Tree C $(\stackrel{\bigcirc}{+})$					
	-	5/12 ^b	7/3	8/26	10/22	5/13	7/4	8/27	10/24	5/14	7/5	8/28	10/24
862	cis-3-Hexenol	0.3	tr ^c	tr	tr	tr	0.9	tr	tr	tr	0.5	0.5	tr
938	α-Thujene	0.5	3.4	5.6	0.2	5.2	4.0	tr	1.3	3.3	3.3	3.1	3.2
944	α-Pinene	0.7	0.9	tr	0.2	1.4	3.8		1.1	tr	tr	tr	tr
980	Sabinene	tr	tr	tr	tr	tr	0.9	tr	tr	tr	0.3	0.9	tr
981	β-Pinene										0.3		
996	Myrcene	0.7	0.9	0.7	0.8	tr	0.7	tr	tr	0.5	0.7	0.7	tr
1026	<i>p</i> -Cymene	0.1											
1031	Limonene	67.4	52.4	56.3	55.8	73.0	59.3	59.9	47.1	63.8	54.2	51.8	43.6
1035	1,8-Cineole	28.4	40.5	34.9	41.0	15.8	26.8	31.9	43.3	12.9	25.4	27.6	35.2
1060	γ-Terpinene	0.6	0.7	0.9	0.9	2.2	1.1	4.2	1.7	2.5	1.7	1.3	2.1
1068	cis-Sabinene hydrate	tr	tr	tr	tr	tr	tr		tr	tr	0.3	0.2	tr
1099	trans-Sabinene hydrate	tr	tr	tr							0.2	0.1	
1103	Linalool	tr	tr	tr		2.5	1.5	2.0	2.2	11.0	6.4	7.8	11.3
1106	Nonanal										0.4	0.5	
1134	cis-Limonene oxide	0.1	tr		tr							tr	
1138	trans-Limonene oxide	0.1	tr	tr	tr	tr	tr		tr	0.3	0.3	0.3	tr
1177	Terpinen-4-ol						tr		tr	tr	0.2	0.2	tr
1192	a-Terpineol	0.7	1.1	1.6	0.9	tr	0.5	tr	0.7	2.7	3.2	3.5	3.5
1216	cis-Sabinene hydrate acetate	0.3	0.2	tr	0.3	tr	tr	0.8	0.6	1.5	0.9	0.7	1.1
1350	α-Terpinyl acetate					tr	0.3	1.2	2.0	tr	tr	tr	tr
1417	β-Caryophyllene	tr	tr	tr	tr	tr	0.2	tr	tr	0.6	0.5	0.4	tr
1475	γ-Muurolene									0.4	0.4	0.2	tr
1493	Valencene									tr	0.2	tr	tr
1512	γ-Cadinene									tr	0.2	tr	tr
1522	δ-Cadinene									0.4	0.4	0.2	tr

^aRI = "Retention Index" determined with reference to a homologous series of *n*-alkanes on an HP-5ms column. ^bDate collected/extracted (day/month). ^ctr = "trace".

generally believed that production of secondary metabolites is energetically costly [19e,20].

Z. clava-herculis is known to be a host plant for the giant swallowtail butterfly, Papilio cresphontes [3]. Other host plants for *P. cresphontes* include *Amvris* elemifera. Ptelea trifoliata, Casimiroa edulis. Zanthoxylum fagara, and Citrus spp. [3,21]. To our knowledge, it is not currently known what phytochemicals may serve as attractants for P. cresphontes to Z. clava-herculis or other members of the Rutaceae. A. elemifera leaf essential oil is rich in limonene and linalool [22] and these two monoterpenoids are abundant in Citrus leaf oils as well Although linalool has been found in many [8,23]. Zanthoxylum spp. [7], limonene was not detected in Z. fagara [24a]. Similarly, P. trifoliata leaf oil showed the presence of linalool, but no limonene [24b]. An analysis of C. edulis leaf oil has shown it to contain neither linalool nor limonene [25a]. Studies are ongoing in our laboratories to determine if leaf essential oils play a role in the attraction of swallowtail butterflies to Z. clavaherculis and other Rutaceae plants.

Experimental

Plant material: A sampling of mature leaves from different locations on three different trees growing in second growth forest at 187 m elevation on the western foot of Green Mountain, Huntsville, Alabama (34° 38' 46" N, 86° 33' 27" W) were collected at four different times of the year (mid-May, early July, late August, and late October) during 2004. The leaves were collected in the morning (approximately 8:00 am), immediately chopped, and hydrodistilled for 4 h using a Likens-Nickerson hydrodistillation-extraction apparatus. Essential oil yields are summarized in Table 1.

Gas chromatographic/mass spectral analysis: The leaf oils of the *Zanthoxylum clava-herculis* samples were subjected to GC-MS analysis as described previously [25a]. The components [25b] from the GC-MS analyses are listed in Table 3.

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